Curriculum Vitae

Dr. David Dulin

Date of birth : 25^{th} of February 1982 ; Citizenship : French.
Family status : in partnership, two children.
Tenured Assistant Professor, UD1
d.dulin@vu.nl
Tel : +31 20 59 83440
Office
Faculty of Sciences/Division of Physics and Astronomy
Vrije Universiteit Amsterdam
De Boelelaan 1081
1081 HV, Amsterdam, The Netherlands
Webpage : https://daviddulinlab.com

Five most important academic achievements

• Established independently the first single-molecule biophysics lab at FAU Erlangen-Nuremberg to investigate protein-nucleic acids interactions, including fully operational molecular biology and microscopy labs.
• Pioneered high-throughput and high-resolution magnetic tweezers set-ups for single-molecule biophysics investigations.
• Pioneered the study of SARS-CoV-2 replication-transcription complex at the single-molecule level, leading to the discovery of a mechanism of action of Remdesivir (first FDA approved antiviral drug against COVID19).
• Established new protocols to fabricate custom-designed high quality nucleic acids construct for single-molecule biophysics assays.
• Developed novel bespoke force and fluorescence spectroscopy microscopes, such as ultra-stable magnetic tweezers and high-throughput TIRF set-ups for single-molecule experiments.

Education

2006-2010	**PhD in Physics** *Paris-Sud XI University, Laboratory Charles Fabry of the Institut d'Optique, France* Specialization in experimental biophysics. Thesis : *"Observation of the translational activity of single ribosomes with fluorescence microscopy coupled to a microfluidic chip"*. Advisor : Prof. Nathalie Westbrook
2000-2006	**Bachelor and Master** Major : physics ; Minor : mathematics , *University of Bordeaux, France*
Languages	French (Native), English (Fluent, C1), German (Basics, B1.1), Dutch (Basics, A2)

Employment

Since 2023	**Tenured Assistant Professor** at *Vrije Universiteit Amsterdam, The Netherlands.*
Since 2021	**Assistant Professor** Physics Department at *Vrije Universiteit Amsterdam, The Netherlands.*
2016-2022	**Junior Research Group Leader non-tenure track** "Physics and Medicine" *IZKF-FAU Erlangen-Nuremberg, Germany*
2014-2016	**Post-doc** in the lab of Achillefs Kapanidis, *Department of Physics, University of Oxford, UK* I established a single-molecule FRET assay to investigate bacterial transcription initiation using TIRF microscopy.
2009-2014	**Post-doc** in the lab of Nynke Dekker, *Department of Bionanosciences, TU Delft, The Netherlands* I pionneered high-throughput and high resolution magnetic tweezers to establish the first assay to investigate elongation dynamics and antiviral drug incorporation of a (+)RNA virus RNA polymerase.
2006-2009	**Research Assistant** in the lab of Nathalie Westbrook, *Laboratory Charles Fabry of the Institut d'Optique, Prof. Alain Aspect lab.* I was the first PhD student in the group establishing a single-molecule biophysics lab, i.e. new custom instrument to build (optical tweezers and TIRF microscope), new biochemical assay and analysis tools.

Funding

Third-party funding (Total : €1,933.5k+US$2,490k)

2022 **Dutch Research Council (NWO) OCENW.M.21.184** Open Competition Domain Science - M, 4 years funding
Revealing how MDA5 interrogates RNA to signal viral infection and trigger innate immunity.
Role : Lead PI.
€359k, including 1 PhD position, consumables and equipment. Will start in March 2023.

2022 **Dutch Research Council (NWO) OCENW.XL21.XL21.115** Open Competition Domain Science - XL, 5 years funding
Know your enemy : deciphering coronavirus biochemical cycles from RNA synthesis to assembly.
Consortium of 8 PIs, with a total funding of €3,032k. Role : Lead PI.
€800k for Dr. Dulin, including 1 PhD, 1 Postdoc and 1 technician positions, consumables and equipment.
Started in January 2023.

2022 **National Institute of Health (NIH), NIAID U19 AI171421** Antiviral Drug Discovery (AViDD), 5 years funding
Rapidly Emerging Antiviral Drug Development Initiative - AViDD Center. Role : co-applicant.
US$945k, including two positions and consumables. Started in May 2022.

2022 **National Institute of Health (NIH), NIAID U19 AI171292** Antiviral Drug Discovery (AViDD), 5 years funding
Development of Outpatient Antiviral Cocktails against SARS-CoV-2 and other Potential Pandemic RNA Viruses. Role : co-applicant.
US$945k, including two positions and consumables. Started in May 2022.

2021 **National Institute of Health (NIH), NIAID R01 AI161841-01** R01, 5 years funding
Coronavirus replication. Role : co-applicant.
US$600k, including 5 years postdoc salary and consumables. Started in March 2021.

2020 **Deutsche Forschungsgemeinschaft (DFG) DU1872/3-1** Individual Research Grant, 2.5 years funding
Revealing the mechanism of directional transcription termination at the single molecule level for the human mitochondrial transcription complex. Role : lead PI.
€276.6k, including 2.5 years postdoc salary and consumables. Started in March 2020.

2020 **Deutsche Forschungsgemeinschaft (DFG) DU1872/5-1** Individual Research Grant, 2.5 years funding
Determinants and dynamics of RNA polymerase I transcription initiation. Role : lead PI.
€276.3k, including 2.5 years postdoc salary and consumables. Started in April 2021.

2020 **Deutsche Forschungsgemeinschaft (DFG) DU1872/4-1** Individual Research Grant, 2 years funding
Revealing the mechanism of nucleotide selection, addition and proofreading of the SARS-coronavirus-1 replication transcription complex at the single molecule level. Role : lead PI.
€221.6k, including 2 years postdoc salary and consumables. Started in August 2021.

Start-up package

2021 **VU Amsterdam - Assistant Professor position**, 5 years position financed by the BaSyC consortium grant, including salary and €400k funding (personnel and material).

2016 **FAU Hospital Erlangen-Nuremberg IZKF Junior Group Leader non-tenure track position** 6 years funded position, including €200k equipment, €300k consumables and €1,076 k personnel.

Professional Service

Academic Service	
2022	Member of the Program committee for the NWO Biophysics 2023 conference.
Service at University	
Since 2021	Member of the Diversity, Equity, and Inclusion committee at the VU Physics and Astronomy Department.
Since 2021	Member of the Examination board of the B.Sc. and M.Sc. "Physics and Astronomy" program of VU Amsterdam and UvA.
2020-2021	Member of the FAU-IZKF Junior committee.

Grant Reviewer Agence Nationale pour la Recherche (ANR), European Research Council (ERC), German Research foundation (DFG).
Peer Reviewer for Nature Structural and Molecular Biology, Nature Communications, Angewandte Chemie International Edition, Nucleic Acids Research, Molecular Cell, Methods, FEBS letter, Journal of Biological Chemistry.

Award

Erasmus exchange fellowship, 2003-2004, *University of Bristol, UK*
ATIP-Avenir CNRS-INSERM, 2020, Competitive funding program from CNRS-INSERM to establish an independant lab in France. Short-listed for the interview ; *declined*.

Teaching related activities

Qualifications	
2023	Supervising PhD Candidates training program, VU Amsterdam.
2023	Basiskwalificatie Onderwijs (BKO, Dutch University Teaching Qualification).
Teaching	**At VU Amsterdam**
Since 2022	Mechanics and Thermodynamics in the Cell. B.Sc (3rd year), *VU and UvA*. Lecturer.
Since 2021	Advanced Biophysics, M.Sc., *VU and UvA*. Lecturer.
	Previously
2018-2020	Magnetic tweezers, *B.Sc. Integrated Life Science, FAU Erlangen-Nuremberg*, lectures and practicum.
2010	Micro and Nanofabrication for biophysics, *Nanosciences M.Sc., TU Delft*, lecture.
2008-2009	Ray optics, microscopy, wave optics for third year B.Sc. at *Institut d'Optique Graduate School*, France. Teaching assistant, practicum.

Publications

Total citations : 1345 ; H-index : 20. Source : Google Scholar

Publications, *preprint*

D. Dulin, Z. Yu, T.J. Cui, B.A. Berghuis, M. Depken and N.H. Dekker
Real-time observation of replicative helicase assembly onto single-stranded DNA
BioRxiv https://doi.org/10.1101/077800, (2016).

H. Dohnalova, M. Seifert, E. Matouskova, F. S. Papini, J. Lipfert, D. Dulin*, F. Lankas*
Real-time observation of replicative helicase assembly onto single-stranded DNA
* : corresponding authors
BioRxiv https://doi.org/10.1101/2023.05.31.543084, (2023).

Publications, *book chapters*

32. D. Dulin
An introduction to magnetic tweezers
In : Heller I., Dulin D., Peterman E. (eds) Single Molecule Analysis : Methods and Protocols, Third Edition. Springer Nature, New York, NY, (2023).
In press

31. S. Quack and D. Dulin
Surface functionalization, nucleic acid tether characterization, and force calibration for a magnetic tweezers assay
In : Heller I., Dulin D., Peterman E. (eds) Single Molecule Analysis : Methods and Protocols, Third Edition. Springer Nature, New York, NY, (2023).
In press

30. E. Ostrofet, F.S. Papini, A.M. Malinen and D. Dulin
A single-molecule view on cellular and viral RNA synthesis
In : Joo C., Rueda D. (eds) Biophysics of RNA-Protein Interactions. Biological and Medical Physics, Biomedical Engineering. Springer, New York, NY, (2019).
https://doi.org/10.1007/978-1-4939-9726-8, (2019).

Publications, *in peer-reviewed journals*

29. R. Chinthapatla, M. Sotoudegan, T. Anderson, I.M. Moustafa, K.T. Passow, S.A. Kennelly, R. Moorthy, D. Dulin, J.Y. Feng, D.A. Harki, R. Kirchdoerfer, C.E. Cameron, J.J. Arnold
Interfering with nucleotide excision by the coronavirus 3'-to-5' exoribonuclease
Nucleic Acids Research, gkac1177 (2022).

28. S.C. Bera, P.P.B. America, S. Maatsola, M. Seifert, E. Ostrofet, J. Cnossen, M. Spermann, F.S. Papini, M. Depken, A.M. Malinen and D. Dulin
Quantitative parameters of bacterial RNA polymerase open-complex formation, stabilization and disruption on a consensus promoter
Nucleic Acids Research, gkac560 (2022).

27. A.M. Malinen, J. Bakermans, E. Aalto-Setala, M. Blessing, D.L.V. Bauer, O. Parilova, G.A. Belogurov, D. Dulin, and A.N. Kapanidis
Real-Time Single-Molecule Studies of RNA Polymerase-Promoter Open Complex Formation Reveal Substantial Heterogeneity Along the Promoter-Opening Pathway
Jounal of Molecular Biology, 434, 2, p.167383 (2021).

26. M. Seifert, S.C. Bera, P. van Nies, R.N. Kirchdoerfer, A. Shannon, T.T.N. Le, X. Meng, H. Xia, J. M. Wood, L. D. Harris, F.S. Papini, J.J. Arnold, S.C. Almo, T.L. Grove, P.-Y. Shi, Y. Xiang, B. Canard, M. Depken, C.E. Cameron, and D. Dulin
Inhibition of SARS-CoV-2 polymerase by nucleotide analogs : a single molecule perspective
eLife, 10 :e70968 (2021)

25. S.C. Bera, M. Seifert, R.N.Kirchdoerfer, P. van Nies, Y. Wubulikasimu, S. Quack, F.S. Papini, J.J. Arnold, B. Canard, C.E. Cameron, M. Depken and D. Dulin
The nucleotide addition cycle of the SARS-CoV-2 polymerase
Cell Reports, 36 (10), 109650 (2021)

24. E. Ostrofet, F. S. Papini, D. Dulin
Microscopy-spectroscopy SI : High spatiotemporal resolution data from a custom magnetic tweezers instrument
Data in Brief, 105397, (2020).

23. M. Seifert, P. van Nies, F.S. Papini, J.J. Arnold, M.M. Poranen, C.E. Cameron, M. Depken and D. Dulin
Temperature controlled high-throughput magnetic tweezers show striking difference in activation energies of replicating viral RNA-dependent RNA polymerases.
Nucleic Acids Research, 48 (10), 5591-5602 (2020)

22. F. S. Papini, M. Seifert and D. Dulin
High-yield fabrication of nucleic acid constructs for single-molecule force and torque spectroscopy experiments.
Nucleic Acids Research, gkz851, (2019).

21. E. Ostrofet, F. S. Papini, D. Dulin
Correction-free force calibration for magnetic tweezers
Scientific Reports, 8,15920, (2018).

20. D. Dulin*, D.L.V. Bauer, A.M. Malinen, J.J.W. Bakermans, M. Kaller, Z. Morichaud, I. Petushkov, M. Depken, K. Brodolin, A. Kulbachinskiy and A.N. Kapanidis*
Pausing controls branching between productive and non-productive pathways during initial transcription in bacteria
Nature Communications, 9 (1), 1478 (2018). **(* : corresponding authors)**.

19. D. Dulin, J.J. Arnold, T. van Laar, H.-S. Oh, C. Lee, D.A. Harki, M. Depken, C.E. Cameron, and N.H. Dekker
Signatures of Nucleotide Analogue Incorporation by an RNA-Dependent RNA Polymerase Revealed Using High-Throughput Magnetic Tweezers
Cell Reports, 21 (4), 1063, (2017).

18. O. Bugaud, N. Barbier, H. Chommy, N. Fiszman, A. Le Gall, D. Dulin, M. Saguy, N. Westbrook, K. Perronet and O. Namy
Kinetics of CrPV and HCV IRES-mediated eukaryotic translation using single-molecule fluorescence microscopy
RNA, 23, 1626, (2017).

17. F. Kriegel, N. Ermann, R. Forbes, D. Dulin, N. H. Dekker and J. Lipfert
Probing the salt dependence of the torsional stiffness of DNA by multiplexed magnetic torque tweezers
Nucl. Acids Res., 45 (10), 5920, (2017).

16. N. N. Vtyurina, D. Dulin, M. Docter, A. Meyer, N.H. Dekker and E. A. Abbondanzieri
Hysteresis in DNA compaction by Dps is described by an Ising model
Proc. Natl. Acad. Sci. U.S.A., 113, 4982, (2016).

15. D. Dulin*, T.J. Cui, J. P. Cnossen, M. W. Docter, J. Lipfert and N.H. Dekker
High Spatiotemporal Resolution Magnetic Tweezers : Calibration and Applications to DNA Dynamics
Biophys. J., 109, 2113, (2015). **(* : corresponding author)**

14. D. Dulin, I. D. Vilfan, B. A. Berghuis, M. Poranen, M. Depken and N.H. Dekker
Backtracking behavior in viral RNA-dependent RNA polymerase provides the basis for a second initiation site
Nucl. Acids Res., 43 (21), 10421, (2015).

13. D. Dulin, B. A. Berghuis, M. Depken and N.H. Dekker
Untangling reaction pathways through modern approaches to high-throughput single-molecule force-spectroscopy experiments
Curr. Op. Struct. Biol., 34, 116, (2015).

12. B.A. Berghuis, D. Dulin, Z.-Q. Xu, T. van Laar, B. Cross, R. Janissen, S. Jergic, N. Dixon, M. Depken and N.H. Dekker
Strand separation suffices to establish a long-lived, foolproof DNA-protein lock at the Tus-Ter replication fork barrier
Nature Chem. Biol., 11, 579, (2015).

11. M.M. van Oene, L.E. Dickinson, F. Pedaci, M. Kober, D. Dulin, J. Lipfert, and N.H. Dekker
Biological magnetometry : Torque on superparamagnetic beads in magnetic fields
Phys. Rev. Lett., 114, 218301, (2015).

10. D. Dulin, I.D. Vilfan, B.A. Berghuis, S. Hage, D. Bamford, M. Poranen, M. Depken, and N.H. Dekker
Elongation-competent pauses govern the fidelity of a viral RNA-dependent RNA polymerase
Cell Reports, 10, 983, (2015).

9. Z. Yu, D. Dulin, J. P. Cnossen, M. Koeber, M. van Oene, O. Ordu, B. A. Berghuis, T. Hensgens, J. Lipfert and N.H. Dekker
A force calibration standard for magnetic tweezers
Rev. Sci. Inst., 85, 123114, (2014).

8. J. P. Cnossen, D. Dulin and N.H. Dekker
An optimized software framework for real-time, high-throughput tracking of spherical beads
Rev. Sci. Inst., 85, 103712, (2014).

7. J. Lipfert, G. M. Skinner, J. M. Keegstra, T. Hensgens, T. Jager, D. Dulin, M. Koeber, Z. Yu, S. P. Donkers, F.-C. Chou, R. Das, and N. H. Dekker
Double-Stranded RNA under Force and Torque : Similarities to and Striking Differences from Double- Stranded DNA
Proc. Natl. Acad. Sci. U.S.A., 111, 15408, (2014).

6. D. Dulin, S. Barland, X. Hachair and F. Pedaci
Efficient illumination for microsecond tracking microscopy
PLoS One, 9, e107335, (2014).

5. R. Janissen, B.A. Berghuis, D. Dulin, M. Wink, T. van Laar and N.H. Dekker
Invincible DNA tethers : covalent DNA anchoring for enhanced temporal and force stability in magnetic tweezers experiments
Nucl. Acids Res., 42, e137, (2014).

4. D. Dulin, J. Lipfert, C. M. Moolman, and N. H. Dekker
Studying genomic processes at the single-molecule level : introducing the tools and applications
Nature Rev. Gen., 14, 9, (2013).

3. A. Le Gall*, D. Dulin*, G. Clavier, R. Meallet-Renault, P. Bouyer, K. Perronet, and N. Westbrook
Improved Photon Yield from a Green Dye with a Reducing and Oxidizing System
Chem. Phys. Chem., 12 (9), 1657, (2011) **(* : equal contribution)**.

2. A. Le Gall, K. Perronet, D. Dulin, A. Villing, P. Bouyer, K. Visscher and N. Westbrook
Simultaneous calibration of optical tweezers spring constant and position detector response
Opt. Exp.,18 (25), 26469, (2010).

Peer-reviewed proceedings

1. D. Dulin, A. Le Gall, K. Perronet, N. Soler, D. Fourmy, S. Yoshizawa, P. Bouyer and N. Westbrook
Reduced photobleaching of BODIPY-FL
Proceedings of HBSM 2009, Physics Procedia, 3 (4), 1563, (2010).

Talks at conferences

* indicates since Principal Investigator

22*. GRC Enzymes, Co-Enzymes and Metabolism (Invited Speaker), *Waterville Valley, USA* (2022)
21*. International Conference on Antiviral Research (ICAR) (Invited Speaker), *Seattle, USA* (2022)
20*. Single-Molecule Biophysics Les Houches *Les Houches, France* (2022)
19*. Dutch Annual Virology Symposium (Invited Speaker), *Amsterdam, The Netherlands* (2021)
18*. CSHL COVID/SARS-CoV-2 Rapid Research Report (Invited Speaker), *Cold Spring Harbour, USA* (2020)
17*. Journees Francophone de la Virologie (Invited Speaker), *Lyon, France* (2019)
16*. Structure and Dynamics of Biomolecules (Invited Speaker), *Huenfeld, Germany* (2019)
15*. Single Molecule Biophysics, *Aspen, USA* (2019)
14*. GFV2018 Annual Meeting of the Society for Virology, *Wuerzburg, Germany* (2018)
13*. SFB960 Symposium The Biology of RNA-Protein Complexes, *Regensburg, Germany* (2017)
12*. FASEB Mechanism and Regulation of Procaryotic Transcription, *Saxons Rivers, USA* (2017)
11. Gordon Research Conference Viruses and Cells, *Girona, Spain* (2015)
10. Society for General Microbiology Annual Conference, *Birmingham, UK* (2015)
9. Biophysical Society Meeting, *San Francisco, USA* (2014)
8. Chemistry in Relation to Biology and Medical Research, *Veldhoven, The Netherlands* (2013)
7. Biophysical Society Meeting, *Philadelphia, USA* (2013)
6. Annual Dutch meeting on Molecular and Cellular Biophysics, *Veldhoven, The Netherlands* (2012)
5. Congress of the French Physical Society, *Bordeaux, France* (2011)
4. Harden Conference RNAP,*Cambridge, England* (2010)
3. Zurich/Paris young scientist meeting, *Paris, France* (2010)
2. Photonics For Life meeting, *Brussels, Belgium* (2008)
1. Congress of the French Physical Society, *Grenoble, France* (2007)

Posters at conferences

10. Single Molecule Biophysics, *Aspen, USA* (2023)
9. Biophysical Society Meeting, *San Diego (USA)* (2020)
8. Single Molecule Biophysics Alpine, *Les Houches (France)* (2020)
7. GRC single molecule, *Mount Snow Resort West Dover, VT (USA)* (2018)
6. GRC single molecule, *Lucca (Barga), Italy* (2014)
5. Harden conference : Machines on genes II, *Oxford (England)* (2012)
4. GRC single molecule, *Mount Snow Resort West Dover, VT (USA)* (2012)
3. Biophysical Society Meeting, *San Diego (USA)* (2012)
2. Annual Dutch meeting on Molecular and Cellular Biophysics, *Veldhoven, The Netherlands* (2010)
1. Biophysical Society Meeting, *Boston (USA)* (2009)

Invited Seminars

* indicates since Principal Investigator

39*. Bionanoscience Department at TU Delft, *Delft (NL)* (2023)
38*. LMS-MRC Imperial College, *London (UK)* (2023)
37*. Department of Microbiology and Immunology, *Chapel Hill (USA)* (2022)
36*. MPI Physics of Light, *Erlangen (Germany)* (2022)
35*. Laboratoire d'Ondes et Matiere d'Aquitaine, *Bordeaux (France)* (2022)
34*. Chair of Biophysics, Utrecht University Physics Department, *Utretcht (The Netherlands)* (2021)
33*. Gene Center, Department of Biochemistry, Ludwig Maximilians University, *Munich (Germany)* (2021)
32*. IZNF, FAU Erlangen *Erlangen (Germany)* (2021)

31*. Laboratoire Architecture et Fonction des Macromolecules Biologiques (AFMB), *Marseille (France)* (2019)
30*. Institute for the Biology and Chemistry of Proteins, *Lyon* (France) (2019)
29*. Structural Biology and Biophysics Seminar, Basel Biozentrum, *Basel* (Switzerland) (2019)
28*. Muenchner Physik Kolloquium, Ludwig Maximilians University, *Munich (Germany)* (2019)
27*. Physikalisches Kolloquium, FAU Erlangen-Nuremberg, *Erlangen (Germany)* (2018)
26*. Bayern Biophotonics, Max Planck Institute Physics of Light, *Erlangen (Germany)* (2017)
25*. GRK1962, FAU Erlangen-Nuremberg, *Erlangen (Germany)* (2017)
24*. Department of Virology, FAU Erlangen-Nuremberg, *Erlangen (Germany)* (2017)
23*. Laboratoire Architecture et Fonction des Macromolecules Biologiques (AFMB), *Marseille (France)* (2017)
22*. Department of Immune Modulation, FAU Erlangen-Nuremberg, *Erlangen (Germany)* (2016)
21. Interdisciplinary Center for Clinical Research (IZKF), FAU Erlangen-Nuremberg, *Erlangen (Germany)* (2015)
20. King's College, *London (UK)* (2015)
19. Centre d'Etudes d'agents Pathogenes et Biotechnologies pour la Sante, *Montpellier (France)* (2015)
18. DIIID Seminar Series, King's College, *London (UK)* (2015)
17. Centre de Biochimie Structurale, *Montpellier (France)* (2015)
16. University of Warwick, *Coventry (UK)* (2015)
15. University of Oxford, *Oxford (UK)* (2014)
14. University of Wollongong, *Wollongong (Australia)* (2013)
13. Centre de Biochimie Structurale, *Montpellier (France)* (2013)
12. Laboratoire Architecture et Fonction des Macromolecules Biologiques (AFMB), *Marseille (France)* (2013)
11. Physics-Biology interface seminar, Laboratoire de Physique Statistique, *Orsay (France)* (2013)
10. Laboratoire de Virologie Moleculaire et Structurale, Gif/Yvette (France) (2012)
9. KNAW Biophysics Meeting, *Amsterdam (The Netherlands)* (2012)
8. Institut Jacques Monod, *University Paris 7, Paris (France)* (2012)
7. Centre de Genetique Moleculaire, *Gif/Yvette (France)* (2012)
6. Laboratoire Charles Fabry of the Institut d'Optique, *Palaiseau (France)* (2010)
5. Centre de Genetique Moleculaire, *Gif/Yvette (France)* (2010)
4. TU Delft, *Delft (The Netherlands)* (2009)
3. Amherst University, *Amherst Massachussets (USA)* (2009)
2. Columbia University, *New-York City (USA)* (2009)
1. Cornell University, *New-York City (USA)* (2009)

Mentorship

Service on PhD examination committees at external institutions

Jetty van Ginkel, TU Delft (The Netherlands)
Olena Parilova, University of Turku (Finland)
Emil Aalto-Setala, University of Turku (Finland)
Paul Poudevigne-Durance, Imperial College (UK)

Supervision since PI

Vrije Universiteit Amsterdam (The Netherlands)
PhD candidates Pim America, Nico van der Vis, Asif Rakib, Arnab Das, Luca Buccolieri.
Postdoctoral fellows Misha Klein, Salina Quack, Sadegh Feiz, Jelmer Cnossen.
Master Student Daniel Buc
Research technician Quinte Smitskamp
Student assistant Eline Bogers

Interdisciplinary Center for Clinical Research (IZKF), FAU Hospital Erlangen-Nuremberg (Germany)
PhD candidates Dr. Mona Seifert (graduated Summa Cum Laude in April 2022), Eugeniu Ostrofet (thesis in preparation)
Research Assistants Dr. Flavia Stal-Papini, Monika Spermann, Yibulayin Wubulikasimu
Postdoctoral Fellows Subhas Chandra Bera, Salina Quack, Sadegh Feiz

Supervision during Postdocs

Department of Physics, University of Oxford (UK)
PhD candidate Rebecca Andrews
Master Students Martin Kaller, Jacob Bakermans

Department of Bionanosciences, TU Delft (The Netherlands)
Postdoctoral fellow Dr. Zhongbo Yu
PhD candidates Natalia Vtyurina, Bojk A. Berghuis, Aartjan te Velthuis
Master Students Tao Ju Cui, Ruadrith Forbes, Jelmer Cnossen, Ivana Cvijovic, Cristina Sfiligoj, Tom Sassen, Matthew Pierotti, Sumit Sachdeva
Bachelor Student Kevin Esajas